TGFβ: A player on multiple fronts in the tumor microenvironment

Fabian Caja¹ and Luca Vannucci¹²

¹Laboratory of Immunotherapy, Institute of Microbiology v.v.i., AS CR, Prague, Czech Republic and ²Laboratory of Tumor Biology, Institute of Animal Physiology and Genetics v.v.i., AS CR, Libechov, Czech Republic

Abstract

The physiological functions of transforming growth factor (TGF)-β in cell signaling include regulation of developmental processes and cell growth. Tumor cells very often display altered regulation of the TGFβ signaling pathway, either by defects in TGFβ itself or in downstream components of the pathway. TGFβ can play a dual role in tumorigenesis, i.e. it can be either tumor-suppressive or tumor-promoting. TGFβ suppresses the growth of tumor cells; however, in advanced tumors, it is associated with induction of progression, resulting in poor prognosis for patients. The TGFβ negative regulation of cytotoxic cell function, together with the promotion of T-regulatory cell maturation, impairs anti-tumor responses. Recent studies have elucidated new roles for TGFβ signaling in the tumor microenvironment. Abrogation of proper signaling induces epithelial-to-mesenchymal transition with pro-metastatic functions, resulting in cancer progression. Thus, TGFβ signaling in the tumor microenvironment plays an important role in tumor initiation, progression, and metastasis by its capacity to regulate cross-talk between tumor cells and other components of the local environment.

Introduction

The malignant changes in healthy cells are sustained and accompanied by alteration of stromal cells and related fibrous structures, forming together the tumor stroma. The tumor cells together with the surrounding immune cells, cancer-associated fibroblasts (CAF), extracellular matrix (ECM) components, blood and lymphatic vessels, and nerves constitute the tumor microenvironment (TM). Many studies proved that the stromal component of TM plays not only a supportive but also a crucial role in cancer development: its components have the capacity to influence and even to deregulate the signaling pathways and interactions between normal and transformed cells in a continuous cross-talk. During embryogenesis, interactions between epithelial and mesenchymal cells in their local environment are essential for the development of tissues and the whole organism. However, in cancer, signaling pathways regulating these interactions are very often deregulated.

Transforming growth factor (TGF)-β, interacting in the TM, is considered as a critical regulator of tumor initiation and progression. TGFβ regulates processes supporting cancer invasiveness, regulation of immune cells of various types, activation, and chemotaxis of fibroblasts. An important mechanism favoring tumorigenesis is the induction of mesenchymal phenotype in the epithelial tumor cells, commonly known as an epithelial-to-mesenchymal transition (EMT). It was proved that this process is induced by prolonged exposition to TGFβ (Miettinen et al., 1994). By this observation, it was suggested that TGFβ plays a dual role in the carcinogenesis. In early phases, TGFβ attenuates proliferation of the tumor cells by activation of growth arrest and apoptosis, but, in advanced tumors, TGFβ promotes tumor cells to be more aggressive and prone to achieve metastatic phenotype (Thiery et al., 2009). TGFβ also suppresses immune response of non-malignant cells and immune cells against cancer through its impact on their differentiation, proliferation, and survival (Li et al., 2006). It promotes angiogenesis and recruits immune cells producing cytokines that stimulate tumor progression (Turner et al., 1990; Wiseman et al., 1988). There are various experimental studies describing the role of TGFβ in initiation of cancer, but more precise investigations of its functions in the TM are still needed. The aim of this review is to address the role of TGFβ signaling in the regulation of TM and, particularly, how it contributes to the progression of cancer. Understanding the critical roles of TGFβ within the TM may provide new targets for design of therapeutics against cancer.

Basic principles of TGFβ signaling

We distinguish three TGFβ molecules: TGFβ1, TGFβ2, and TGFβ3. They are secreted as inactive homodimers and belong to the TGFβ protein superfamily. This includes 33 members in humans, such as activins, inhibins, bone morphogenic protein (BMP), growth and differentiation factors (GDF), glial cell line-derived neurotrophic factor (BDNF), and the above-mentioned TGFβ protein members (Massague, 2012; Piek et al., 1999). The most abundant isoform is TGFβ1, a 44-kDa protein, coded by TGFBI gene located at chromosome 19 (chromosome 7 in mice). It is ubiquitously expressed in all tissues (Derynck et al., 1985). TGFβ2 is a 48-kDa protein coded by TGFBI gene located at chromosome 1. TGFβ2 is expressed in neurons and astroglial cells of embryonic tissues (Flanders et al., 1991), and it effects the
heart, as well as other mesenchymal structures and development. In the adult mouse, it is expressed in almost all tissues, especially in the placenta, the male submaxillary gland and the lung, but not in the liver (Boyer et al., 1999; Miller et al., 1989). The TGF/β2 transcript attenuates T-cell maturation and immune responses in the TM, thus it is supporting tumor growth (Schwyzer & Fontana, 1985). TGF/β3 is a 47-kDa protein coded by TGFβ3 gene located at chromosome 14 (chromosome 12 in mice). It functions as a regulator of palate development, in which it regulates cellular adhesion and formation of ECM (Proetzel et al., 1995), but it is also important for lung development and for wound healing in the skin (Bandyopadhyay et al., 2006; Kaartinen et al., 1995). TGFβ3 is expressed mainly in the umbilical cord (Stewart et al., 1996).

TGF/β molecules, deposited in the extracellular matrix (ECM), interact with three TGF/β receptor types (T/RI, T/RII, and T/RIII). All TGF/β ligands differ in their binding affinity to T/RII. Both TGF/β1 and TGF/β3 bind to T/RII. TGF/β2 needs T/RI as a co-interacting partner for high-affinity interaction with T/RII, which binds alone to T/RIII (Derynck & Zhang, 2003). T/RI and T/RII are predominantly localized at the cell membrane in a homodimeric conformation. After binding TGF/β, T/RII is activated by autophosphorylation and forms a heterotetrameric complex with T/RI. Thereafter, T/RI transphosphorylates and activates T/RI. This mechanism allows T/RI to phosphorylate its downstream mediators SMAD2 and SMAD3 (Shi & Massague, 2003). Two types of TGF/β signaling cascades have been identified (Figure 1).

The canonical one is SMAD-dependent and the non-canonical one is SMAD-independent. In general, the canonical cascade involves phosphorylation of the carboxy-terminal serine residues of the SMAD2 and SMAD3 proteins that are receptor-regulated SMADs (also called receptor-SMADs; R-SMADs). Phosphorylation allows their oligomerization with SMAD4, also known as ‘co-SMAD’. This interaction is necessary for transcription of the complex to the nucleus (Schmierer & Hill, 2005) in order to modulate gene transcription. SMAD 7 competitively inhibits SMAD2/3 binding to T/RI (Inoue & Imamura, 2008).

Non-canonical signaling involves activation of PI3K-AKT, RhoA, Rac1, Ras, Cdc42, Daxx, Par6, TAB1/TAK1, and MAPK pathways (Bierie & Moses, 2006). These pathways are more complex than the canonical one and involve more intensive cross-talk between them. Among them, the Rho-Rock1 and AKT pathways activated by TGF/β significantly contribute to migratory and invasive cellular phenotypes observed in various types of cancer (Dumont et al., 2003). Pleiotropic TGF/β ligands are involved in many other processes; for example, they suppress cell proliferation by repressing CDK4 expression and by activating the expression of CDK inhibitors (Ewen et al., 1995; Polyak et al., 1994). SMAD-dependent activation of Bcl-2 proteins is important for regulation of programmed cell death (Pardali & Moustakas, 2007). In addition, the regulation of cellular adhesions by TGF/β signaling is very important for tumorigenesis, mainly via decreases in E-cadherin and zonula adherens 1 production and through cyto-skeletal re-arrangements (Huber et al., 2005).

Figure 1. Representation of main TGF/β1 downstream pathways related to tumor microenvironment. See also text.
Taken together, the above-mentioned facts highlight the double-edged character of TGFβ signaling.

**TGFβ production and activation**

TGFβ molecules are primarily synthesized as homodimers, stabilized by disulfide bridges and non-covalent interactions, and undergo intracellular processing before they act in signaling cascades (Dubois et al., 1995). First, these pro-proteins are cleaved in trans-Golgi apparatus by furin proteases to release truncated TGFβ dimer and a resting dimeric component called ‘latency-associated protein’ (LAP). Subsequently, LAP interacts with TGFβ to form the ‘small latent complex’ (SLC). Finally, SLC associates with latent TGFβ binding glycoprotein (LTBP) to form a 240kDa large latent complex (LLC) (Miyazono et al., 1988). The LLC is secreted to the ECM network, where it is deposited in an inactive form (Rifkin, 2005). The LTBP protein is necessary for storing TGFβ in the ECM and, thus, plays a key role in TGFβ accumulation and release. The LTBP family includes four LTBP isomers (LTBP1–4) structurally similar to fibrillin. Each LTBP contains two types of cysteine-rich domains, i.e. an eight-cysteine domain and epidermal growth factor (EGF)-like repeats (Rifkin, 2005). All isomers of LTBP contribute to tumorigenesis in various types of cancer.

Many physiological processes and different factors activating extracellularly-deposited TGFβ from latent complex have been described in vivo so far, e.g. retinoic acid, integrins, matrix metalloproteases (MMP)-2, MMP-9, reactive oxygen species (ROS), irradiation, and thrombospondin-1 (TSP-1) (Barcellos-Hoff & Dix, 1996; Munger et al., 1999; Schultz-Cherry & Hoff, 1996; Munger et al., 1999; Schultz-Cherry & Murphy-Ullrich, 1993; Yu & Stamenkovic, 2000). In addition, TGFβ can be activated by a decrease in the pH in a local environment. For example, an acidic environment is formed in vivo by osteoclasts attached to bone tissue during resorption. It was shown in in vitro experiments that the pH in this site was low enough to activate proteases that, in turn, allowed for the release of latent TGFβ complexes (Oursler, 1994). The protease plasmin also has numerous functions in the TGFβ activation cascade in vivo (Lyons et al., 1988). Specifically, LAP is proteolytically cleaved by plasmin, with a change of LTBP complex conformation and release of mature TGFβ from the complex. Retinoic acid can also activate latent TGFβ by similar processes (Kojima & Rifkin, 1993).

Interestingly, TSP-1 not only has an anti-angiogenic role, but it also appears to play a role in cancer initiation and progression through other mechanisms (Lawler & Detmar, 2004). Even mechanical tensions in the ECM can allow release of TGFβ from stored LTBP complex, a mechanism possible under tissue stiffening during chronic inflammation or tumor progression (Wipff et al., 2007; Wipff & Hinz, 2008). Each of the abovementioned factors interfere with the non-covalent interactions between LAP and mature TGFβ and, via this mechanism, they allow TGFβ to be released from its latent state.

**Role of TGFβ in cancer**

It is well known that components of the TGFβ signaling cascade are very often deregulated in various types of cancer. As noted above, TGFβ has a dual role in tumorigenesis; it can be a tumor-suppressing or tumor-promoting factor, depending on the stage of tumor development. Tumor suppression is promoted by repressing expression of c-Myc and cyclin-dependent kinase genes (CDKs) and by activating expression of CDK inhibitor genes p15, p21 and p27 (Datto et al., 1995; Hannon & Beach, 1994; Polyak et al., 1994). TGFβ is also able to down-regulate or inhibit expression of CDK4 and CDC25A genes (Iavarone & Massague, 1999). The second role of TGFβ, as a cancer promoter, is exerted through an inhibition of apoptosis and/or by a stimulation of proliferation.

Normally, TGFβ acts as a tumor suppressor in mature tissues and is generally produced in the TM. How then is it possible that tumor cells can proliferate in such suppressive environment? Cancer cells have evolved many strategies on how to use TGFβ for their survival. Typically, transformed cells can have mutated or disrupted TGFβ receptors or altered SMAD signaling pathways. Especially in breast, prostate, and colorectal carcinoma (CRC), alterations in the TGFβ signaling cascade can have prognostic significance (Bierie & Moses, 2006).

**TGFBR2** is probably the most commonly affected gene from all genes coding components of the cascade. It codes one of the most important proteins of the cascade-TβRII, which recognizes and binds all isomers of TGFβ. Repressed or down-regulated expression of TGFBR2 is found in many types of cancer and it is leading to increased tumor spreading. In addition, it is associated with the microsatellite instability in CRC. Hereditary and sporadic CRC tend to have high microsatellite instability in 10-bp poly-A sequence of TGFBR2, causing malfunction of TβRII (Kim et al., 2000). Apart from APC, K-RAS and TP53 genes, also microsatellite stable CRCs display mutations in TGFBR2. Moreover, TGFBR1, SMAD2, and SMAD4 genes are very commonly lost, mutated, or functionally attenuated. For example, TGFβ1T869C polymorphism is associated with 2.7-fold greater relative risk of developing squamous cell carcinoma, suggesting that also gene polymorphisms can affect the proper functions of TGFβ protein (Carneiro et al., 2013). Still, the real contribution of the gene polymorphisms on the development of various types of cancer still needs to be clarified.

Another type of protein, E3 ligase Smurf2, is commonly up-regulated in squamous cell carcinomas with low levels of SMAD2 phosphorylation (Fukuchi et al., 2002). DNA methylation of TGFBR1 and TGFBR2 genes was observed in some cancers, suggesting the existence of epigenetic mechanisms regulating the pathway (Kang et al., 1999). An increased angiogenesis and invasion is induced by SMAD-independent up-regulation of MMP expression (Safina et al., 2007). Interestingly, TGFβ signaling in the malignant phenotype is able to regulate microRNA (miR) function. For example, hepatocellular carcinoma cells express CC-chemokine ligand 22 (CCL22) only when expression of miR-34a is inhibited by TGFβ (Yang et al., 2012). TGFβ increases the expression of miR-29a, which induces angiogenesis and represses the expression of phosphatase and tensin homolog (PTEN) (Wang et al., 2013). Important too is the TGFβ-induced expression of miR-494 in myeloid-derived suppressor cells (MDSC); this leads to increases in expression of CXC chemokine receptor and reduction in the expression of PTEN. These regulations also lead to increased expression of MMP3, MMP13, and MMP14 (Liu et al., 2012). Further, tumor-associated natural killer (NK) cells are silenced by TGFβ-inducible miR-183 (Donatelli et al., 2014).

The importance of TGFβ signaling in tumorigenesis has been studied in vitro by many investigators, mimicking conditions of tumors in patients by preparing TGFβ gene mutants or by directly treating the cancer cells with TGFβ. For example, Sartor et al. (2010) proved that TGFβ had a capacity to increase expression of genes coding collagen type 1, collagen type 2, MMP2, MMP9, and lysyl oxidase homolog 4 in A549 lung adenocarcinoma cells. Those authors also observed increased expression of vascular endothelial growth factor A (VEGFA) and TSP-1. Advanced tumor stages are characterized by epithelial changes, but also by changes affecting the TM. Elevated TGFβ signaling is associated with increased metastases and poor prognosis for patients. Interestingly, loss of TGFβ signaling also correlates with increased metastases and progression and with poor prognosis.
Deregulation of TGFβ signaling is very often associated with stromal changes, such as activation of fibroblasts, deposition of ECM, increased angiogenesis and infiltration of immune cells (Bierie & Moses, 2006). Finally, in consideration of the heterogeneity of cancer cells in a tumor, it may be supposed that only part of these cells could be sensitive to TGFβ. However, this hypothesis still warrants further investigation. Interestingly, stromal changes induced by altered TGFβ expression were found to increase metastatic activity of TGFβ unresponsive tumor cells (Finak et al., 2008). Thus, TGFβ can induce tumor progression directly or indirectly.

### TGFβ regulation of immune cells

TGFβ is considered one of the most important regulators of proliferation and differentiation of immune cells deposited in a TM (Table 1). TGFβ is produced by and binds to many different types of immune cells, including macrophages, dendritic cells (DC), NK cells, B-cells, and T-cells. Cancer cells can also produce TGFβ; therefore, TGFβ has a capacity to modulate innate as well as adaptive immunity under both physiologic and cancer states (Yang et al., 2010).

In B-cells, TGFβ regulates expression of immunoglobulins, surface receptors, and major histocompatibility complex type II proteins (MHC II). These proteins are the direct markers of B-cell maturation and differentiation (Lebman & Edmiston, 1999). TGFβ also regulates T-cell maturation. It also inhibits proliferation of naïve CD4+ cells and T-cell expansion (Gilbert et al., 1997). Experiments on transgenic mice bearing dominant-negative TGFBR2 gene showed there was spontaneous T-cell differentiation leading to development of autoimmune diseases (Gorelik & Flavell, 2002). TGFβ favors tumor progression by suppressing T-cell production of perforins, granzymes, and other toxins. Thus, TGFβ negatively regulates both the expansion and cytotoxic activity of CD8+ T-cells, functions crucial to anti-tumor immunity (Thomas & Massague, 2005).

#### Table 1. Regulation of immune cell function by TGFβ.

<table>
<thead>
<tr>
<th>Regulated function</th>
<th>Effect of TGFβ</th>
<th>Types of immune cells</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maturation</td>
<td>Up-regulation</td>
<td>DC, Macrophages (MΦ)</td>
</tr>
<tr>
<td>Antigen presentation</td>
<td>Up-regulation</td>
<td>DC, Natural killer cells (NK)</td>
</tr>
<tr>
<td>Chemotaxis</td>
<td>Up-regulation</td>
<td>DC, MO, Mast cells</td>
</tr>
<tr>
<td>Proliferation</td>
<td>Up-regulation</td>
<td>CD8+ T cells, CD4+ T cells, Neutrophils</td>
</tr>
<tr>
<td>Effector function</td>
<td>Up-regulation</td>
<td>CD8+ T cells, CD4+ T cells</td>
</tr>
<tr>
<td>T1H1 and T1H2 cells</td>
<td>Up-regulation</td>
<td>CD4+ T cells, CD8+ T cells</td>
</tr>
<tr>
<td>Treg cells</td>
<td>Up-regulation</td>
<td>CD4+ T cells</td>
</tr>
<tr>
<td>IgA class switching</td>
<td>Inhibition</td>
<td>B cells</td>
</tr>
<tr>
<td>Activation</td>
<td>Inhibition</td>
<td>B cells</td>
</tr>
<tr>
<td>Apoptosis</td>
<td>Inhibition</td>
<td>B cells, NK</td>
</tr>
<tr>
<td>Cytotoxicity</td>
<td>Inhibition</td>
<td>Neutrophils</td>
</tr>
<tr>
<td>Inflammatory cytokine</td>
<td>Up-regulation</td>
<td>MΦ, Neutrophils</td>
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<tr>
<td>secretion</td>
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<tr>
<td>Polarisation from N1 type</td>
<td>Up-regulation</td>
<td>MΦ, Neutrophils</td>
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<td>to N2 type</td>
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<tr>
<td>Polarisation from M1 type</td>
<td>Up-regulation</td>
<td>MΦ, Neutrophils</td>
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<td>to M2 type</td>
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†, Up-regulation; †, inhibition.

TGFβ also has a capacity to induce FoxP3 gene expression and subsequently to generate regulatory T (Treg)-cells. Together with interleukin (IL)-6, TGFβ induces T1H17 cells that produce IL-17, important for activation of leukocytes (Shevach, 2009; Weaver et al., 2006). SMAD4+ T-cells producing Th12-type cytokines that promote stromal expansion were found in gastrointestinal tumors (Kim et al., 2006). TGFβ inhibits the proper maturation of NK cells, which then lose their capacity to recognize non-self antigens, a process important for clearance of tumor cells (Marcoe et al., 2012). Moreover, TGFβ negatively regulates the ability of DC to present foreign antigens (Tanaka et al., 2010).

Proliferation of monocyte–macrophage lineage cells is suppressed mainly by TGFβ ligands (Chantry et al., 1989; Tsunawaki et al., 1988). In the TM, two types of macrophages can be found: M1 that deliver active anti-cancer functions and M2 type that promote tumor progression and metastasis. M2 are the most abundant inside tumors, and are also known as tumor-associated macrophages (TAM) (Mantovani et al., 2006). TGFβ is able to induce a shift of polarization from anti-tumor M1 to M2 TAM (Gong et al., 2012). Interestingly, in vitro inhibition of TGFβ signaling in TAM, blocking the T/R and ligating the toll-like receptor 7 by an agonist reconverted M2 type macrophages into M1 type, provided new perspectives in cancer therapy (Peng et al., 2013). Recently, it was discovered that TGFβ has the capacity to transform anti-tumorigenic neutrophils (N1) into protumorigenic neutrophils (N2) associated with production of MMP9 and chemokine CXCL1 (Fridlender et al., 2009). Interestingly, depletion of TGFβ results in reversible polarization of N2 neutrophils to N1 with an anti-tumor phenotype. Unexpected properties of TGFβ were recently described. TGFβ administered to mesenchymal stem cells (MSC), instead of leading to an increase, reversed their immunosuppressive activity upon T-cells. Moreover, TGFβ produced by MSC was found (in an autocrine manner) to inhibit inflammatory cytokine-induced inducible nitric oxide synthase (iNOS) expressed by MSCs themselves (Xu et al., 2014a).

The most important regulator of inflammation, NF-κB, is also negatively regulated by TGFβ via activation of inhibitor of kappa B (IκB) protein, with down-regulation of the pro-inflammatory and pro-metastatic functions of NF-κB. However, in some studies on cell lines, a possible double-faceted effect of TGFβ was noted, with TGFβ either promoting or inhibiting NF-κB functions (Arsura et al., 1996; Han et al., 1998). A recent study on gastric cancer development showed how TGFβ effects are microenvironmental. In a TGFβ mutant, with impeded binding of TGFβ to the latent TGFβ binding protein (Tgfb1−/−,33), generalized inflammation and increased tumorigenesis developed. By introducing a second mutation into, and thereby subsequent suppression of, the recombination activator gene 2 (Tgfb1−/−,33, Rag2−/−), the inflammation and pro-carcinogenic effects did not appear. Those authors indicated this experiment showed how changes in tumor onset were more directly associated with inflammatory processes, rather than with the loss of TGFβ protein. This experiment also highlights the role of TGFβ in controlling inflammation (Ota et al., 2014).

### TGFβ as a regulator of tumor microenvironment

The tumor microenvironment is very dynamic and the active crosstalk within the various types of involved cells (both cancer and non-cancer cells) permits a tumor to establish and progress, escaping host immunosurveillance and anti-cancer responses. The TM is commonly a hypoxic area with a low pH, conditions supporting DNA damage and suppressed repair (Bristow et al., 2008). Moreover, it is fueled by persistent inflammation that importantly contributes in supporting and promoting tumor...
development and spread (Balkwill & Mantovani, 2001). Normal TGFβ1 master regulation of inflammation in physiological conditions turns to inhibitory and re-modeling functions in the TM, frustrating the anti-cancer response efficacy. Further, the organ microenvironment can affect progression of tumors, as recently described for experimental hepatocellular carcinoma (HCC). For example, when human HCC cells were inoculated in the subcutaneous or in the liver of nu/nu mice, based on the different sites of development, TGFβ1 mRNA levels were found to be significantly lower in liver tumors than in subcutaneous tumors, and these levels correlated with higher tumor weight and less pulmonary metastasis for the orthotopic cancers (Li et al., 2013).

TGFβ was for the first time observed as a regulator of TM when Bhownick et al. (2004) found that deletion of the TGFBR2 gene in mouse fibroblasts was inducing transformation of adjacent prostate and stomach epithelia. In vivo, various epithelial cells also displayed deletion of TGFBR2, and this deletion resulted in increased tumor progression and metastatic growth (Yang & Moses, 2008). Moreover, hepatocyte growth factor (HGF) and HGF receptor MET are very often up-regulated in tissues displaying TGFβ down-regulation, suggesting an important role of paracrine signaling in these tissues. Recently, an experiment was designed in which induction of TGFβ in CAF stimulated production of IL-11, thereby triggering STAT3. Mice treated with TGFBR2 inhibitor were not able to form metastases (Calon et al., 2012).

TGFβ is also involved in regulation of chemokines, chemokine receptors, and angiogenesis. For example, breast cancer cells increasingly produce TGFβ, which induce production of angiopoietin-like 4 proteins, thereby enhancing formation of metastases in lungs (Padua et al., 2008). However, loss of TβRII in these cancer cells correlates with recruitment of F4/80+ cells that produce pro-inflammatory proteins CXCL1 and CXCL5 (Yang & Moses, 2008). This complete loss of TGFβ signaling in epithelial cells correlates with reduced survival in patients with breast cancer, especially estrogen-receptor-positive patients (Bierie et al., 2009). Even the loss of TβRIII contributes to tumor progression. Hanks et al. (2013) elucidated a new mechanism in melanoma and breast cancer cells in which loss of tumor-produced TβRIII induced the production of indoleamine 2,3-dioxigenase in plasmacytoid DC and of CCL22 chemokines in myeloid DC, thereby mediating Treg infiltration and suppression of anti-tumor immunity. Further, hypoxic conditions (a characteristic marker of TM) were seen to promote breast cancer as a result of mesenchymal stem cell secretion of TGFβ (Huang et al., 2013).

Other patterns have been observed in gastric carcinoma and in colon cancer models. In gastric cancer SNU16mAd cells, an SMAD-dependent pathway activates production of integrins through protein kinase Cδ (PKCδ), thereby enhancing invasiveness of the tumor cells (Lee et al., 2005). In cis-Apc+/-Smad3-/mice, an increased recruitment of CCR1+ myeloid cells with promotion of colon cancer cell invasiveness was found (Kitamura et al., 2007). The blockage of TGFβ was found to increase expression of pro-inflammatory cytokine genes such as IL-5, IL-6, and IL-13. While this can lead to the negative effects about promotion of tumor progression described by Mantovani et al. (2006), it also increases the response against tumor elicited by specific immunotherapy (Kim et al., 2006). Once again, the context and timing of cytokine network activity is critical. Increased inflammation in the TM was also observed in various non-GI cancers (e.g. head and neck carcinomas), and this was put in relation to deregulation of TGFβ signaling. Crosstalk between TGFβ and IL-1 signaling pathways appears to be very common in some cancer cell lines (Lu et al., 2007).

Lastly, TGFβ is important for EMT regulation, a key process leading to tumor invasion and metastases formation (Thiery, 2002). EMT occurs during wound healing, normal cell development, and abnormally in cancer progression in which epithelial cells differentiate into mesenchymal cells (Thiery et al., 2009). EMT is associated with transition of primordial epithelial cells during gastrulation, generating neural crest cells and formation of endocardial tissue. Epithelial cells can transform into fibroblasts during wound healing, regeneration, and fibrosis. EMT is also a characteristic process accompanying cancerogenesis (Zeisberg & Neilson, 2009). The EMT implies disruption of tight junctions and delocalization of tight junction proteins, disruption of adherent junctions, and re-organization of actin fibers. Epithelial cells display mesenchymal markers and show spindle-like morphology (Thiery, 2002). TGFβ is responsible for EMT maintenance through production of protein surviving that stabilizes tubulin and Aurora B, resulting in inhibition of cell cycle arrest and apoptosis (Lee et al., 2013). Moreover, colon cancer cells are able to transform normal fibroblasts into CAF by secretion of TGFβ (Hawinkels et al., 2014). CLIC4 (chloride intracellular channel 4) is a downstream effector of the TGFβ signaling pathway, regulating transition of normal fibroblasts to activated pro-metastatic myofibroblasts through p38 signaling. Renal, ovarian, and breast cancers showed increased production of CLIC4, which should be considered as a new target of anti-tumor therapy (Shukla et al., 2013; Suh et al., 2007). TGFβ is also responsible for tumor recurrence through IL-8-dependent activation of cancer stem-like cells, as was shown in patients with breast cancer (Bhola et al., 2013). EMT can also be initiated in epidermal keratinocytes by ROS-stimulated TGFβ secretion and MAPK activation (Fukawa et al., 2012).

**Therapeutic perspectives**

Since TGFβ plays dual roles in tumorigenesis, it would seem to be an intriguing prospective therapeutic target. It was demonstrated in several studies that loss of TGFβ signaling is not tumorigenic but can affect already pro-tumorigenic (inflammatory) micro-environments. Conversely, over-expression of TGBF genes is commonly associated with progression of aggressive tumors with pro-metastatic potential and poor patient prognosis. Many neutralizing antibodies and molecular inhibitors that suppress tumor-promoting functions of TGFβ have been discovered so far. However, it is very important to design drugs that do not affect the normal tumor suppressive properties of TGFβ (Kim et al., 2008).

Several clinical studies proved that TGFβ therapy can be safe and effective (Bogdahn et al., 2011; Schlingensiepen et al., 2011). The main advantages for reducing TGFβ are reported as a better host immune surveillance and better prognosis for patients after radio- or chemotherapy (Biswas et al., 2007). TGFβ modulation also has effects on TM, i.e. it induces T-cell-mediated anti-tumor responses by causing an increased infiltration of NK cells and T-cells into the TM. TGFβ also helps to reduce the suppressive capacity of Treg cells and to decrease production of IL-17 that inhibits apoptosis in tumor cells (Nakamura et al., 2001; Nam et al., 2008). Another study showed that even SMAD4-deficient tumors could be treated by TGFβ therapy, suggesting pro-tumorigenic functions of TGFβ depend on complex TGFβ signaling in the TM (Zhong et al., 2010). Immunotherapy with TGFβ and tumor necrosis factor (TNF)-α antagonists was found to be able to restore production of interferon (IFN)-α by tumor-associated DC, resulting in anti-tumor responses (Sisirak et al., 2013).

MED12, a key component of transcription MEDIATOR system, has become a new target for therapy. MEDIATOR is a protein system that integrates and transduces positive and
negative regulatory information, from enhancers and operators to promoters, functioning through RNA polymerase II with modulation of its activity in promoter-dependent transcription (Myers & Kornberg, 2000). In vitro experiments showed that loss or suppression of MED12 is associated with EMT and drug resistance due to activation of T/JRIL MED12−/− cells treated by TGFβ signaling inhibitors displayed restoration of drug responsiveness (Huang et al., 2012). Xu et al. (2014b) designed an experiment based on nanotechnology principles. Anti-TGFβ small interfering RNA was nanoparticle-delivered into the late-stage TM. This administration down-regulated TGFβ production, leading to enhanced therapeutic effects of a vaccine against melanoma tumors in C57BL/6 mice (Xu et al., 2014b).

Despite all the positive effects of TGFβ modulation, Achyut et al. (2013) discovered that abrogation of TGFβ signaling in stromal cells of Tgfbr2−/−/−KO mice increased expression of various inflammatory mediators (e.g. iNOS and cyclooxygenase 2), inducing genetic damage, and proliferation in the neighboring epithelial compartment. Expression of the downstream mediator of p53, Cdkn1a/p21 (p21), was reduced. This, when taken together with the increases in inflammation and inflammatory cell infiltration, could also enhance tumor progression. The different effects of TGFβ infiltration, could also enhance tumor progression. The different expression and functions of mature cells. TGFβ is also an important factor in the tumorigenesis network. Interestingly, TGFβ plays both tumor-suppressive and -promoting roles. It is evident that cross-talk between different cells in a TM is essential for cancer progression and that TGFβ is a potent regulator of this cross-talk. It regulates tumor progression by mutual interactions between various components of TM, including fibroblasts, epithelial cells, stromal cells, immune cells, etc. An increase in the pro-metastatic capacity of tumor cells is induced through the EMT, yet also regulated by TGFβ signaling. These facts have led to a strong effort to target TGFβ in anti-tumor therapy; various very promising drugs that have been discovered so far. Despite this success, the exact roles of TGFβ in the TM still need further elucidation not only to permit the better design of new therapeutic approaches, but to also more precisely define strategies for intervention.

Declaration of interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

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